

# PSOSP uncovers pervasive SOS-independent prophages with distinct genomic and host traits in the bacterial genomes

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Yali Hao, Mujie Zhang, Xinjuan Lei, Chengrui Zhu, Taoliang Zhang, Yanping Zheng, Xiang Xiao, Huahua Jian. 2025.  
PSOSP uncovers pervasive SOS-independent prophages with distinct genomic and host traits in the bacterial  
genomes. *iMeta* 4: e70073. <https://doi.org/10.1002/imt2.70073>

# Introduction

- the **classical** lysogenic-lytic switch of prophage relies on the **bacterial SOS pathway**
- recently, more **SOS pathway-independent induction** of prophages has been reported
- induction experiments using MMC on environmental microorganisms have shown **highly variable induction rates, with widespread insensitivity**

eg: **237 human intestinal** lysogenic bacterial strains : **1/3** of the prophages can be activated by MMC and H<sub>2</sub>O<sub>2</sub>

(A)

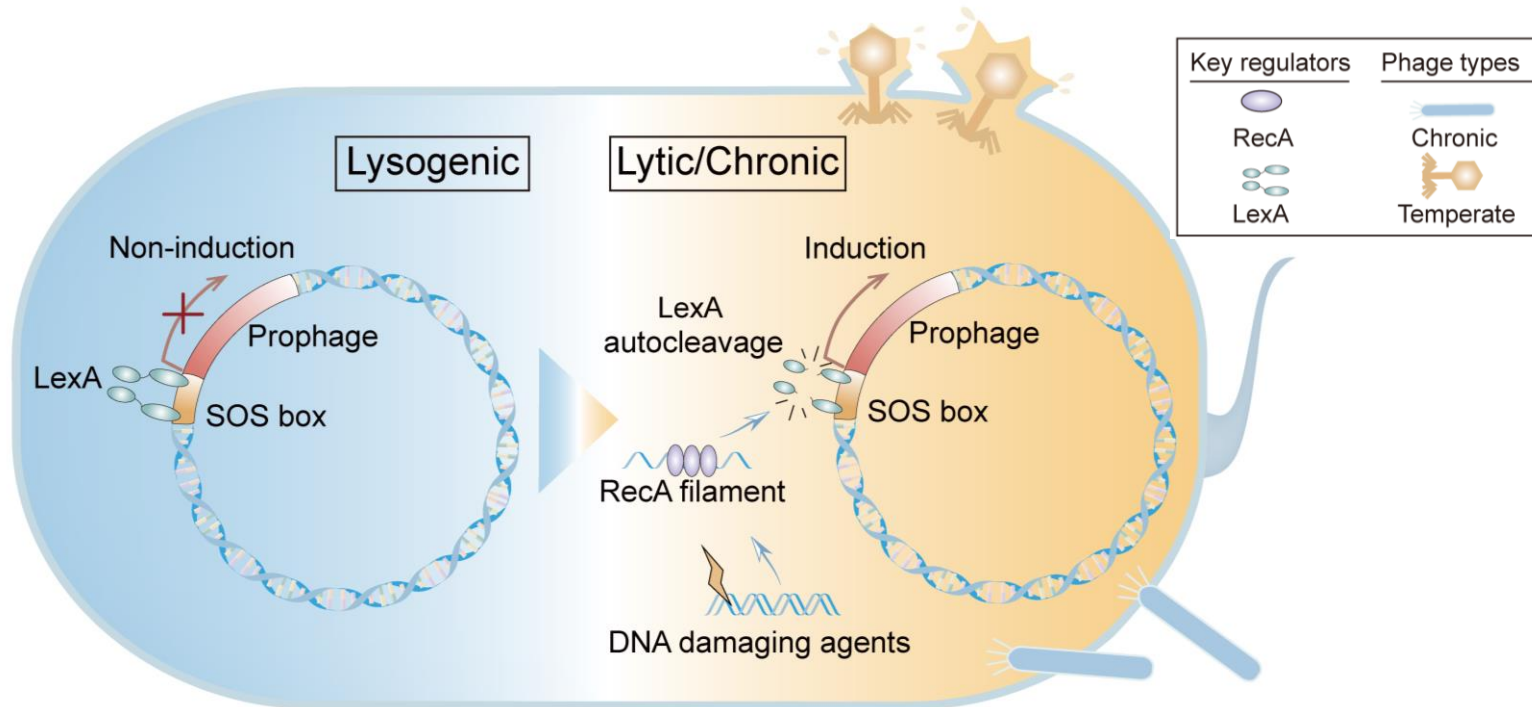
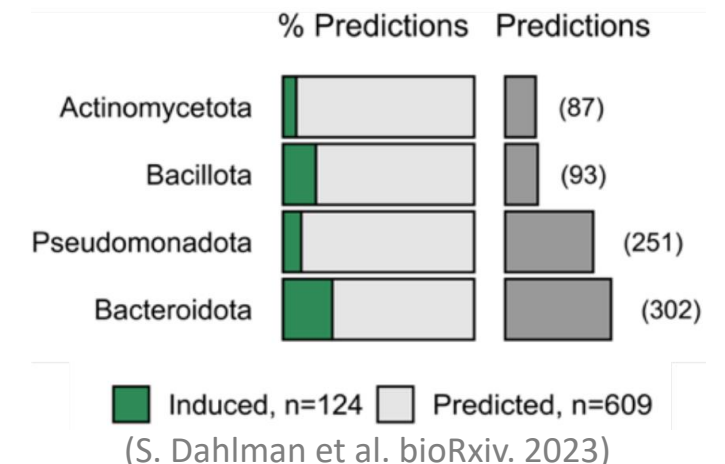
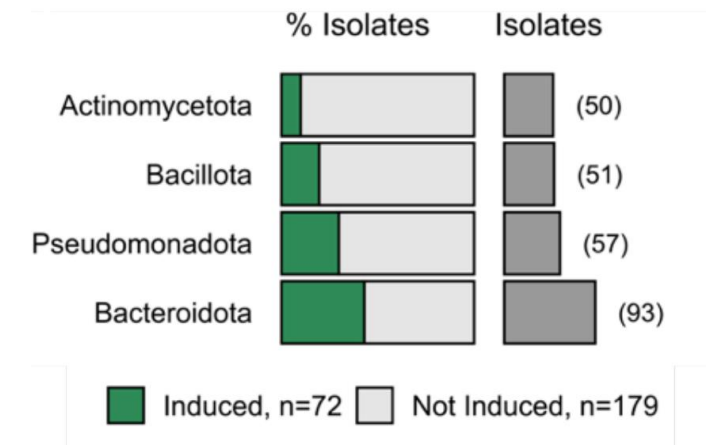
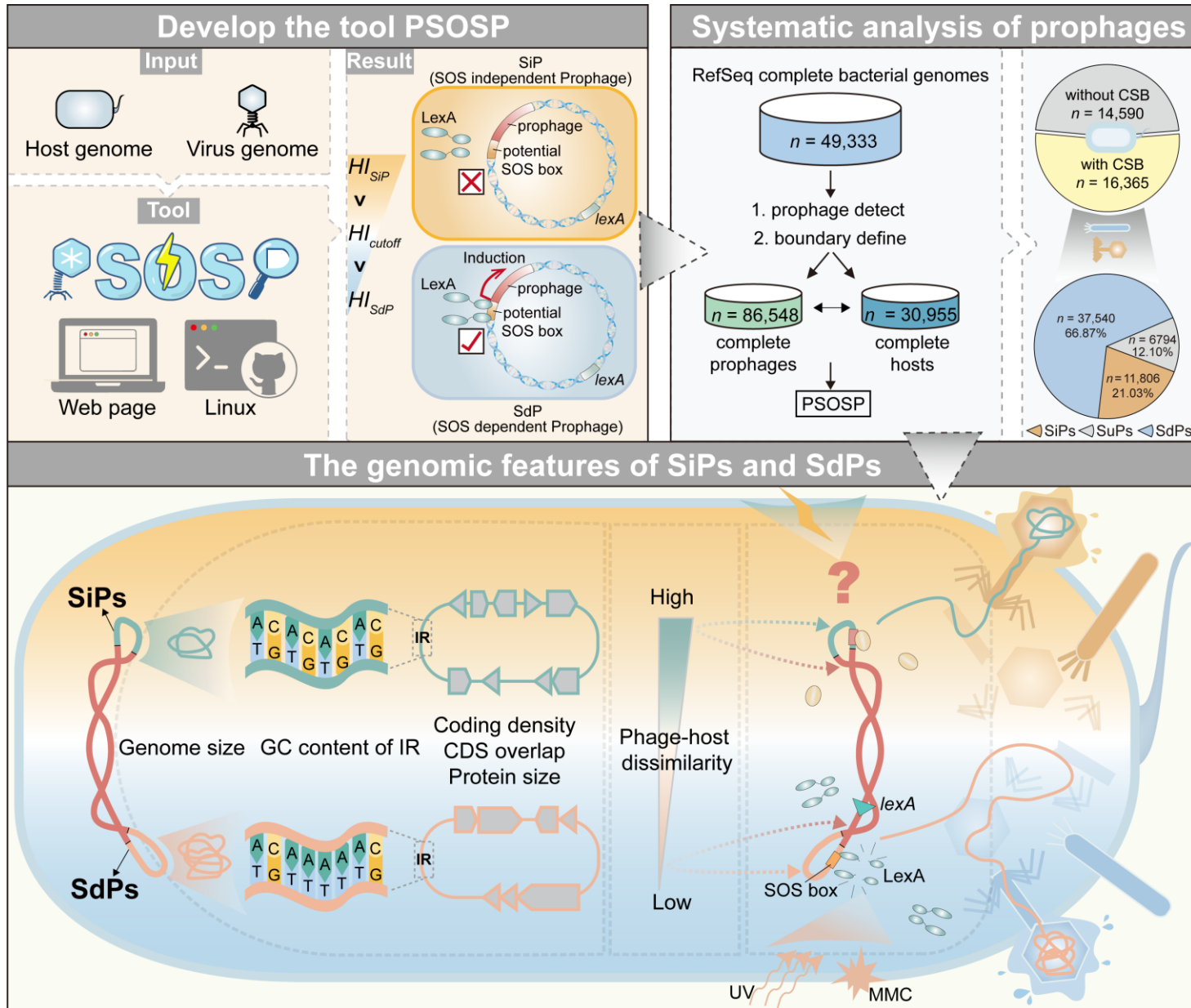


Figure 1(A) Schematic diagram of the currently known lysogenic-lytic switch mechanism in temperate phages.

**Limitation:** due to the lack of specialized tools for determining prophage induction modes, the distribution proportions and genomic characteristics of SOS-independent prophages (SiPs) remain largely unexplored



# Highlights



- Develop a novel bioinformatics tool **PSOSP** that **predicts prophages induction modes**;  
Website: <https://vee-lab.sjtu.edu.cn/PSOSP/>  
Github: <https://github.com/mujiezhang/PSOSP>
- **Identify 11,806 SiPs** by applying PSOSP to 49,333 complete bacterial genomes;
- Uncover that **SiPs and SdPs exhibit distinct genomic and host traits**, suggesting the **potential for mutual conversion** between certain SiP and SdP groups;
- Refine the conventional understanding of **temperate phage induction mechanisms** and **provide novel tools** and insights for exploring the lysogenic-lytic switch of phages

# Result 1: *HI* reliably predicts LexA binding potential

## ● The workflow of PSOSP:

- (1) scanning the host genome to identify LexA protein and canonical SOS boxes (CSBs) located upstream of the *lexA* gene;
  - (2) identifying potential SOS boxes (PSBs) across bacterial genomes, calculating the Heterology Index (*HI*) for each PSB and establishing classification thresholds ( $HI_{C1}$  and  $HI_{C2}$ ) via Mean Shift clustering results;
  - (3) scanning PSBs within prophage promoter regions and determining of the minimum *HI* ( $HI_{min}$ );
  - (4) evaluating the ability of LexA binding to prophage promoter regions by comparing  $HI_{min}$  with thresholds ( $HI_{C1}$  and  $HI_{C2}$ ), and subsequently classifying the induction modes of prophage
- The binding interactions between previously reported LexA protein with **PSBs ( $n = 24$ ) in *E. coli* K12** (as documented by Lewis et al.) could be **precisely predicted based on *HI***

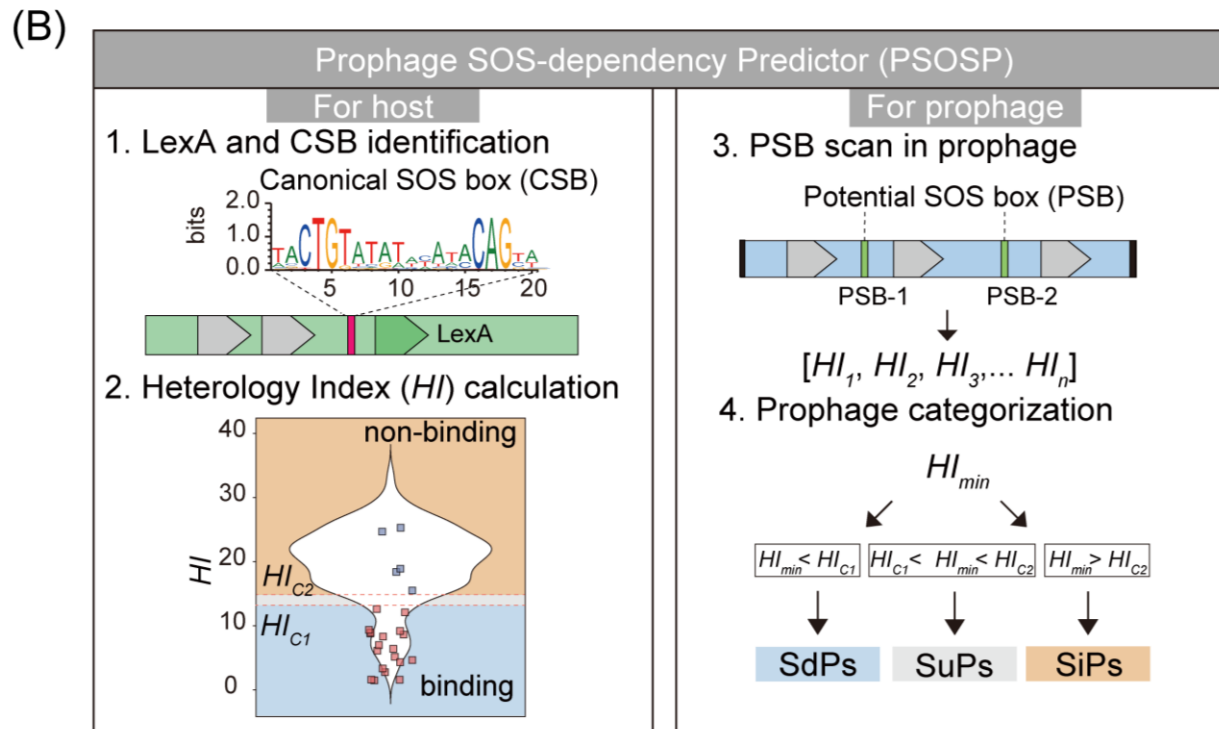


Figure 1(B) Schematic diagram of the PSOSP workflow

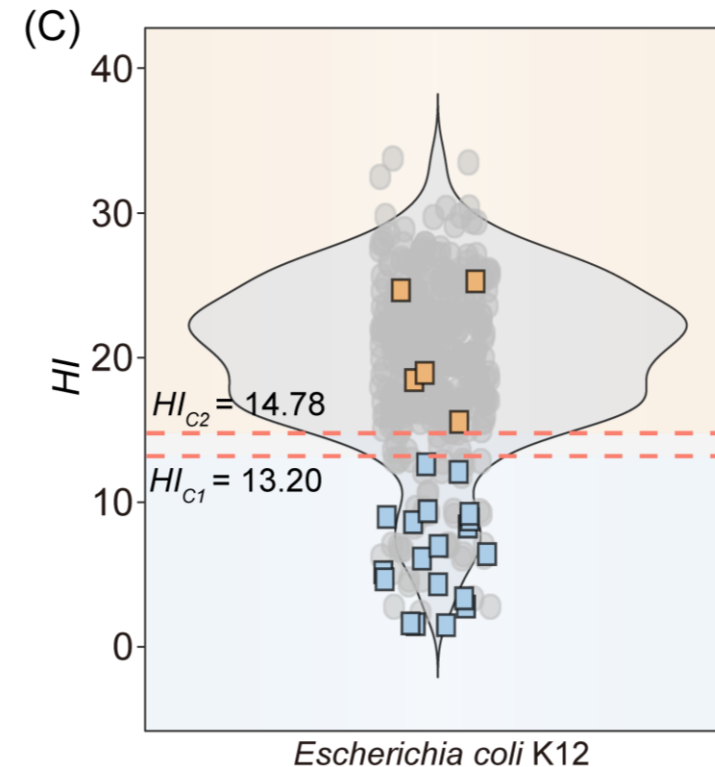


Figure 1(C) Distribution of *HI* for all PSBs in *E. coli* K12



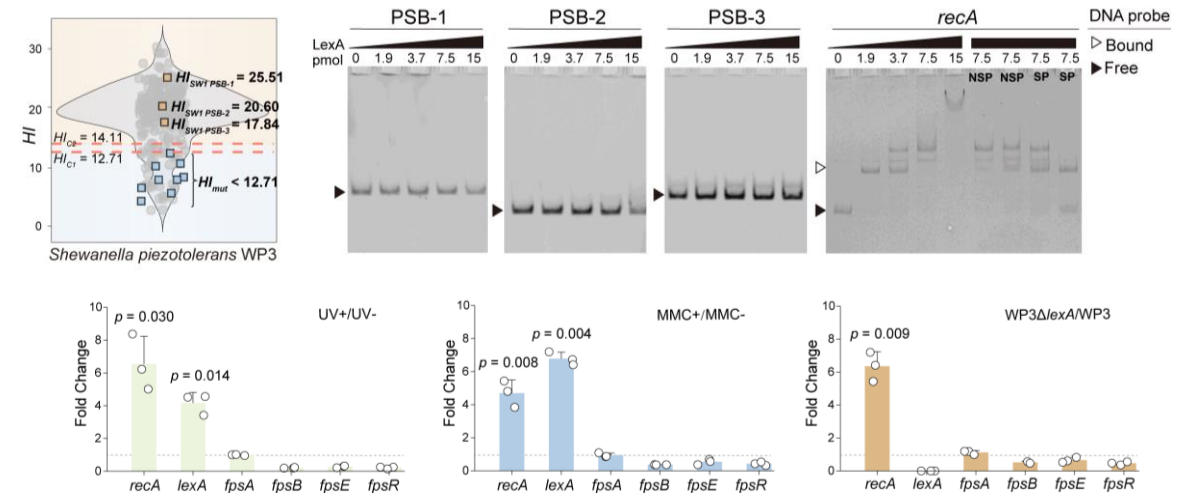
# Result 2: PSOSP: determining the regulatory mode of prophages based on *HI*

**Table S4. Experimentally validated induction-mode bacteriophages and hosts in this study.**

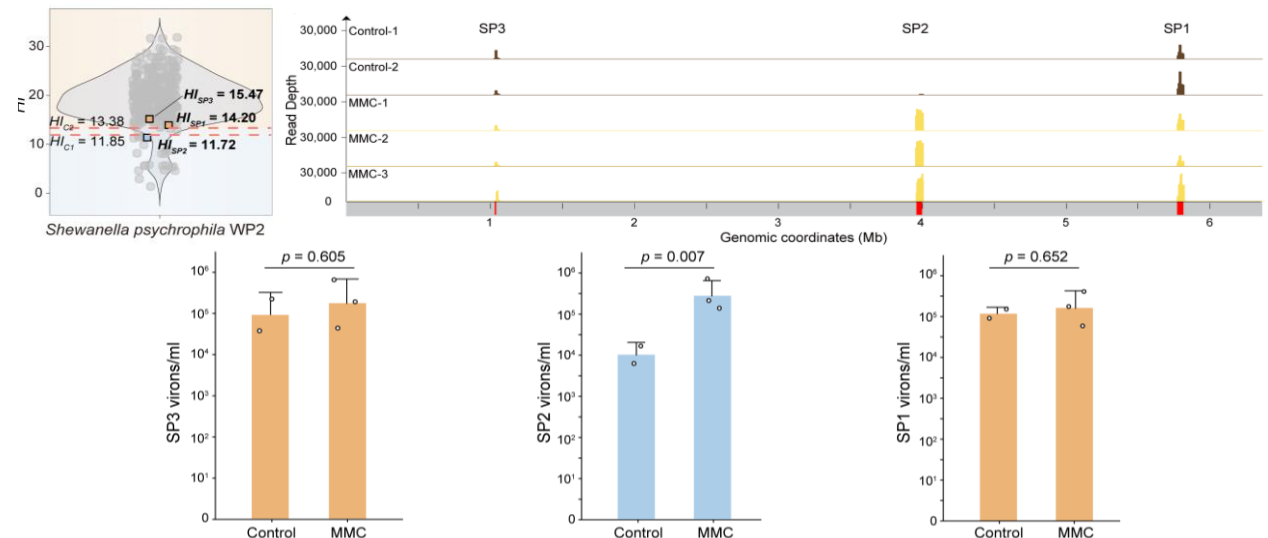
Prophage	Host taxonomy (Genus)	Phage genome size (bp)	Experimentally validated induction mode	PSOSP predicted induction mode
P22	<i>Salmonella</i>	41,724	SdP	SdP
Fels-1	<i>Salmonella</i>	42,723	SdP	SdP
Fels-2	<i>Salmonella</i>	33,693	SdP	SdP
phiECO1	<i>Escherichia</i>	31,478	SdP	SdP
ST-8624	<i>Escherichia</i>	62,822	SdP	SdP
VALGphi6	<i>Vibrio</i>	8,530	SiP	SiP
B3	<i>Pseudomonas</i>	38,439	SiP	SiP
vB_SspS_OS31	<i>Serratia_J</i>	42,280	SdP	SdP
vB_SspM_BZS1	<i>Serratia_J</i>	44,995	SdP	SdP
yong1	<i>Hafnia</i>	43,329	SdP	SdP

- **Test set: ten prophages** (8.5–62.8 kb) belonging to **eight viral taxonomic families**, with hosts spanning **seven different genera**.
- PSOSP achieve **100% sensitivity and 100% specificity** in the test set

## ● The validation of prophage SW1 in *Shewanella piezotolerans* WP3



## ● The validation of prophage SP1, SP2, SP3 in *S. psychrophila* WP2



# Result 3: Systematic analysis of SiPs and SdPs in bacterial genomes

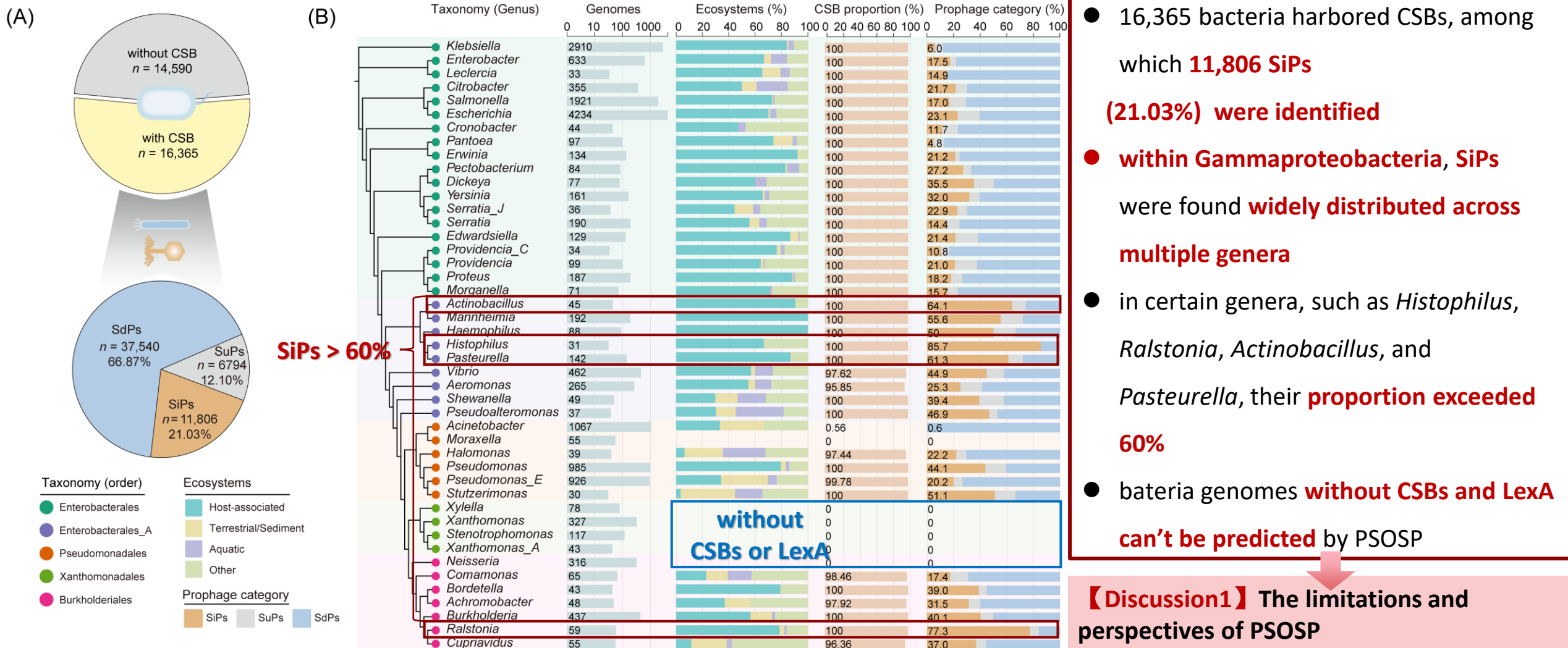
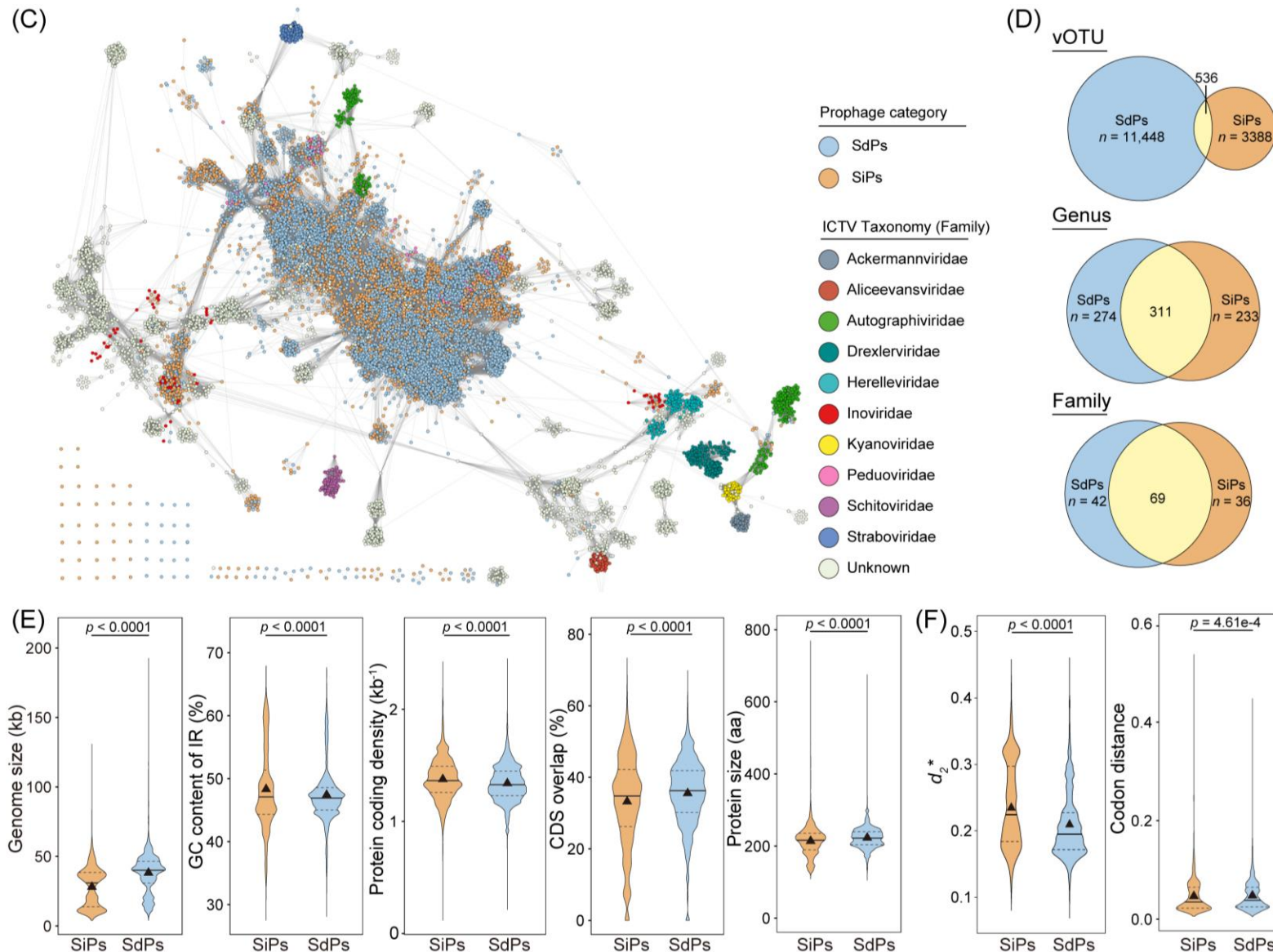


Figure 2. Widespread existence of SiPs and the comparison between SiPs and SdPs

# Result 3: Systematic analysis of SiPs and SdPs in bacterial genomes



- SiPs and SdPs exhibit both distinct **clustering** and **overlapping** patterns
- at different taxonomic levels, **the degree of shared features between SiPs and SdPs increased with higher taxonomic ranks**

【Discussion2】 The clustering feature may suggest **the potential for mutual conversion** between certain SiP and SdP groups

- compared to SdPs, SiPs have a significantly **lower** proportion of CDS overlap (PCO) and smaller protein sizes, but **higher** GC content of intergenic region (IR) and protein-coding density (PCD)
- SiPs exhibited greater nucleotide feature divergence from their hosts** compared to SdPs, suggesting **lower compatibility with their hosts**

Figure 2. Widespread existence of SiPs and the comparison between SiPs and SdPs



# Summary

- ❑ We developed **a novel bioinformatics tool PSOSP** to predict prophage induction modes. PSOSP was **experimentally validated** to accurately distinguish SdPs from SiPs.
- ❑ We discovered that **SiPs were widely distributed within bacterial genomes and exhibited distinct genomic features** compared to the more well-studied SdPs. Correspondingly, **the hosts** of these two prophage types are hypothesized to **differ in their physiological characteristics**.
- ❑ These PSOSP-enabled findings provide not only novel insights into **diverse induction mechanisms** but also **a critical methodology** for future studies on phage-host interactions and prophage isolation strategies.
- ❑ PSOSP website: <https://vee-lab.sjtu.edu.cn/PSOSP/>  
PSOSP Github: <https://github.com/mujiezhang/PSOSP>

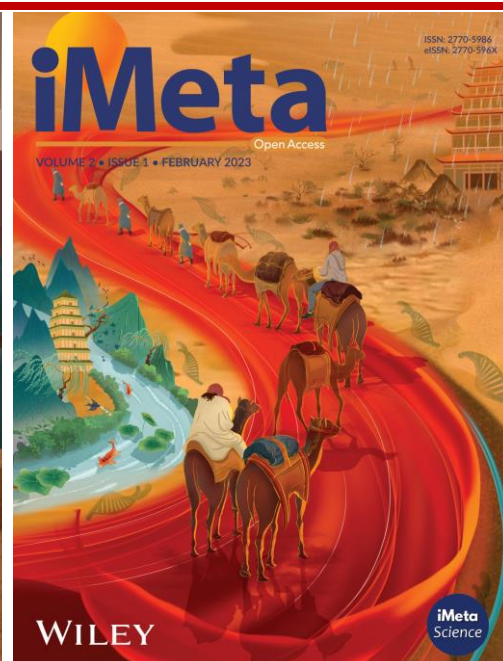


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