



Species-level exploration of the gut microbiome in the leaf-eating Presbytis monkeys reflected the effects of anthropogenic activity and specialized dietary niches: conservation on the fourth biodiversity level

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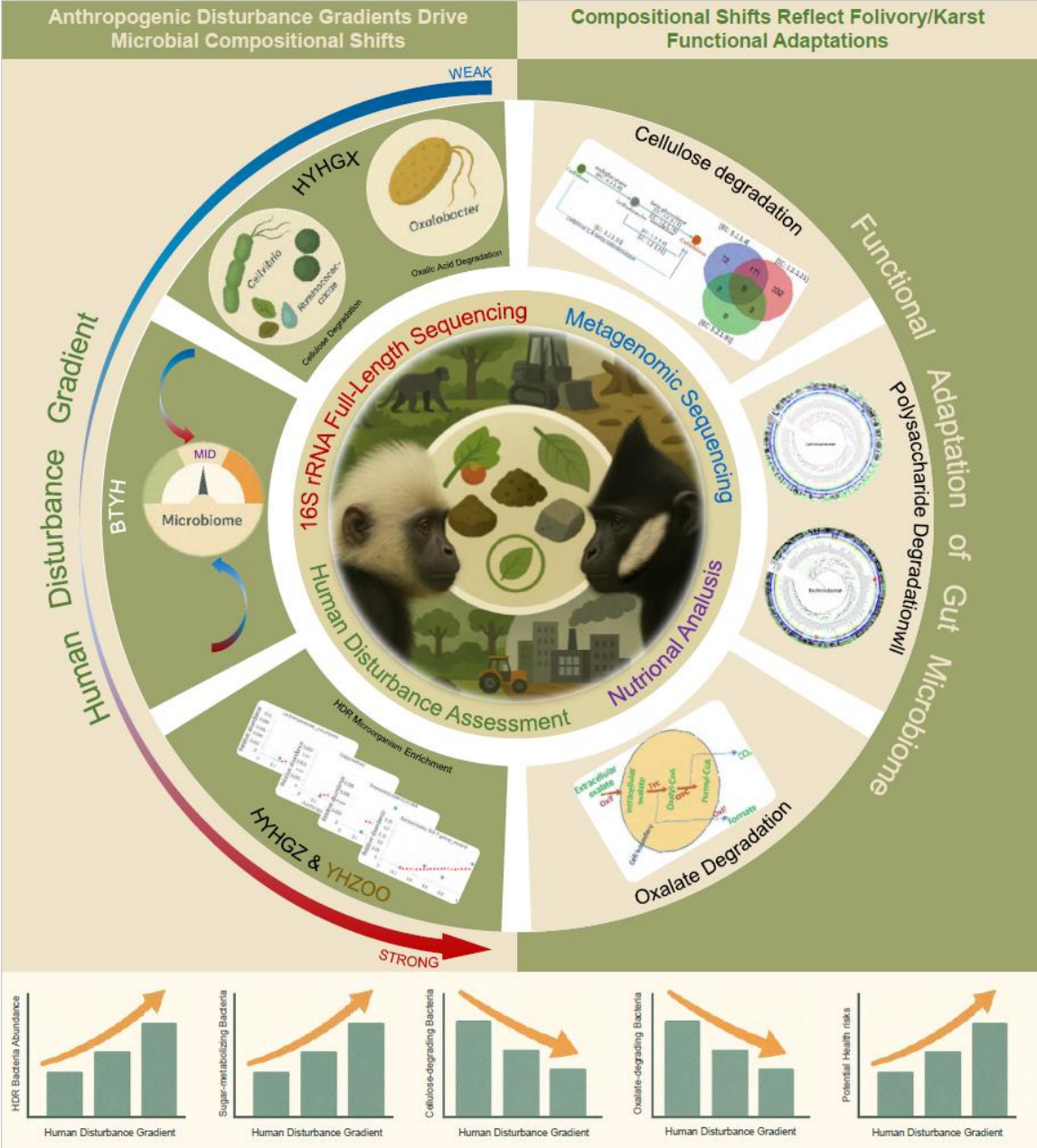
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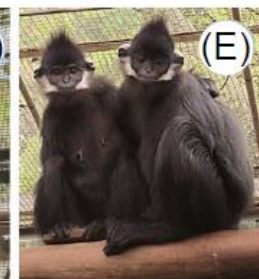
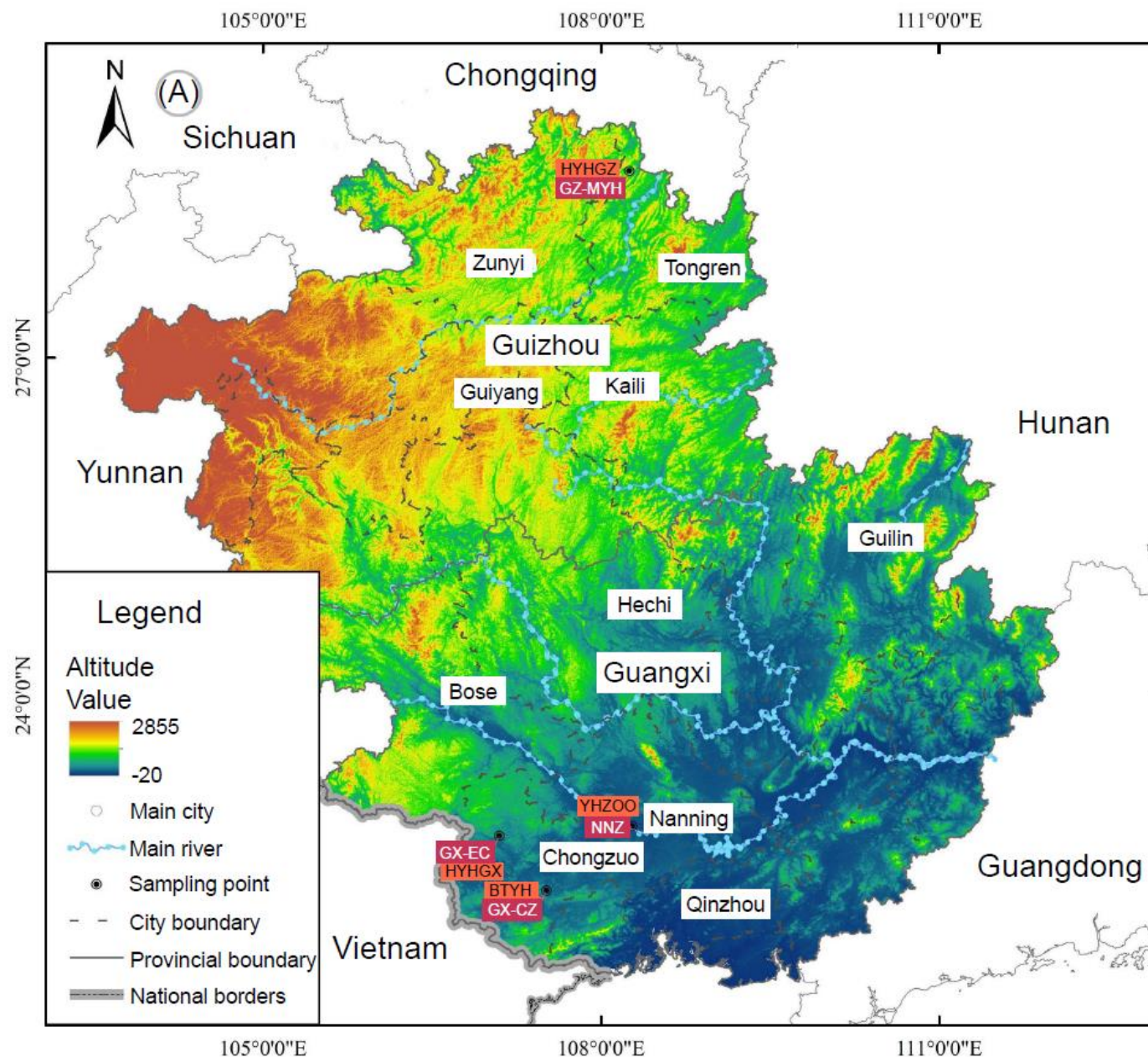


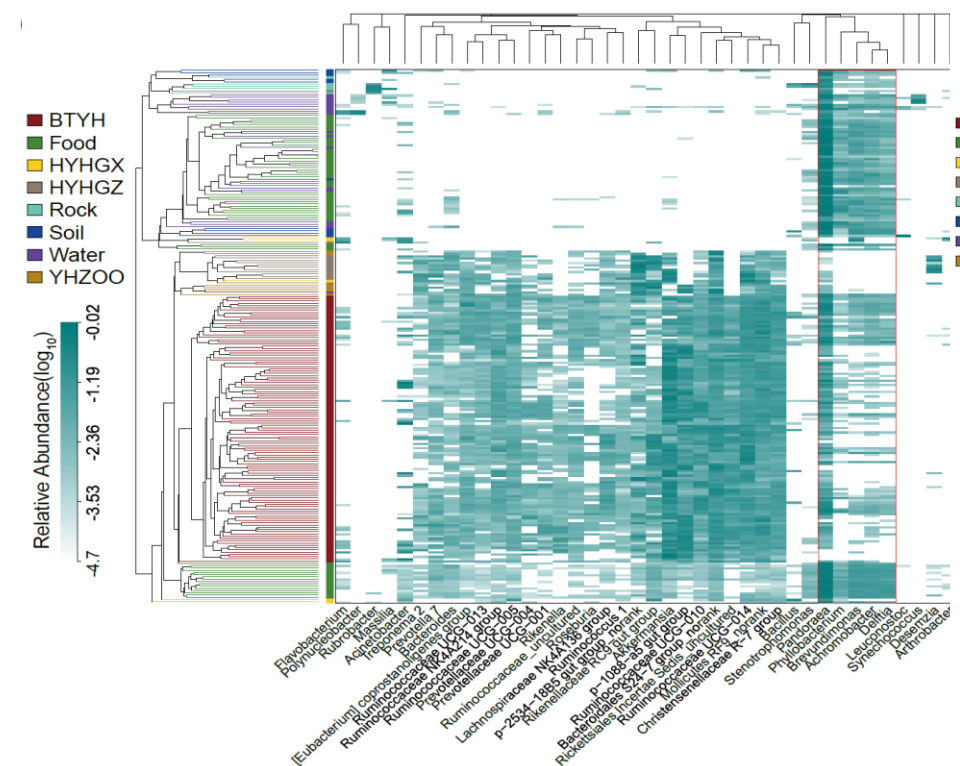
Qihai Zhou, Qiuyan Guo, Xinyuan Cui, Tao Meng, Zhuting Pang, Song Wang, Hua Chen, et al. 2025. Species-level exploration of the gut microbiome in the leaf-eating Presbytis monkeys reflected the effects of anthropogenic activity and specialized dietary niches: conservation on the fourth biodiversity level. *iMetaOmics* 2: e70051. <https://doi.org/10.1002/imo2.70051>



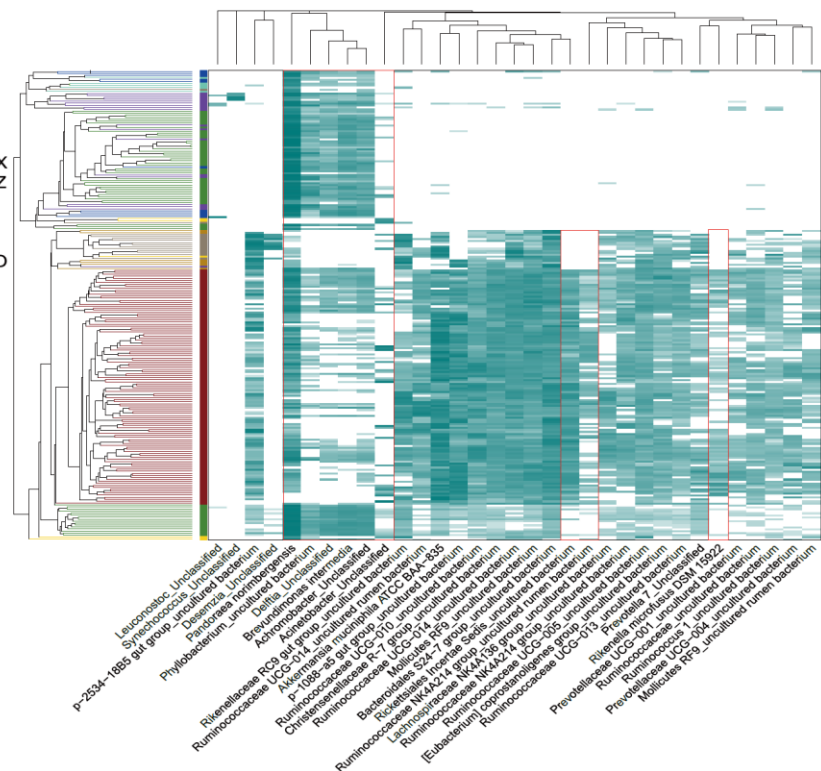


Highlights

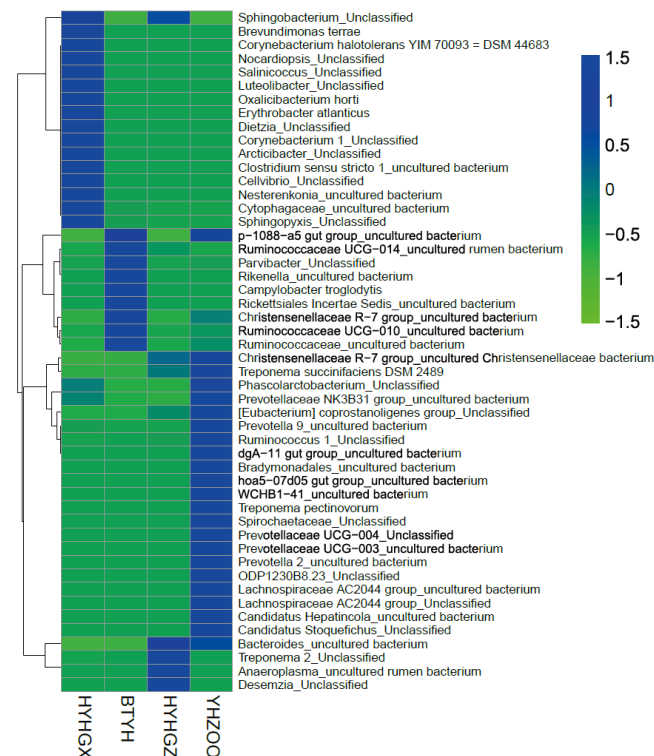




Dominant Genera Composition

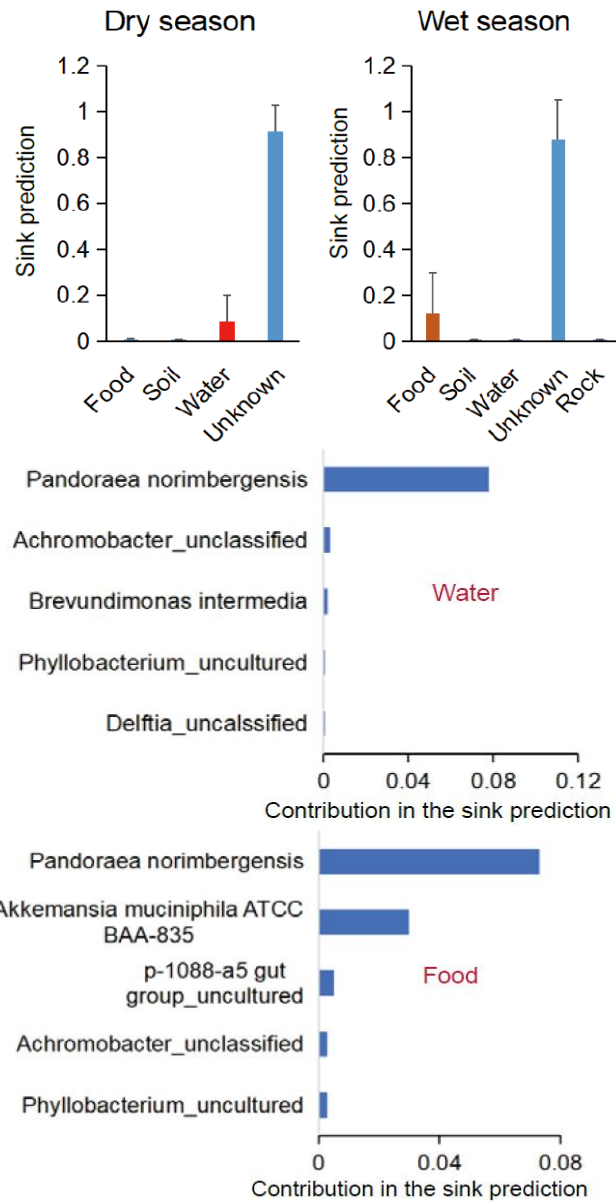
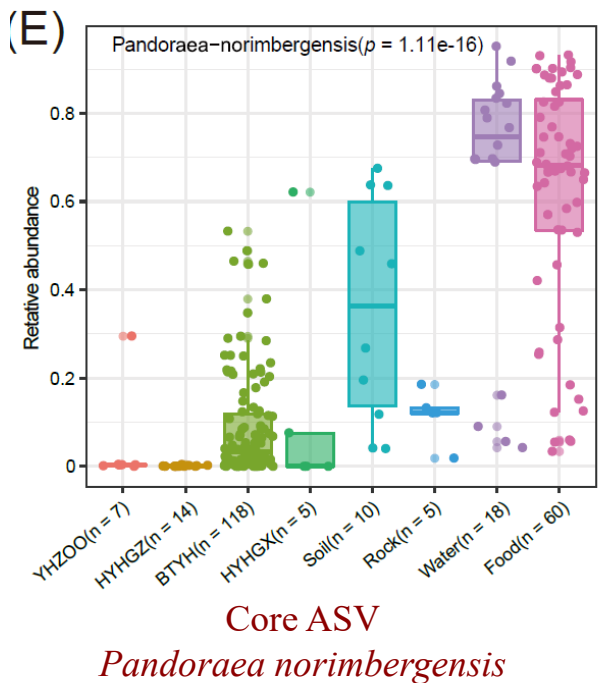
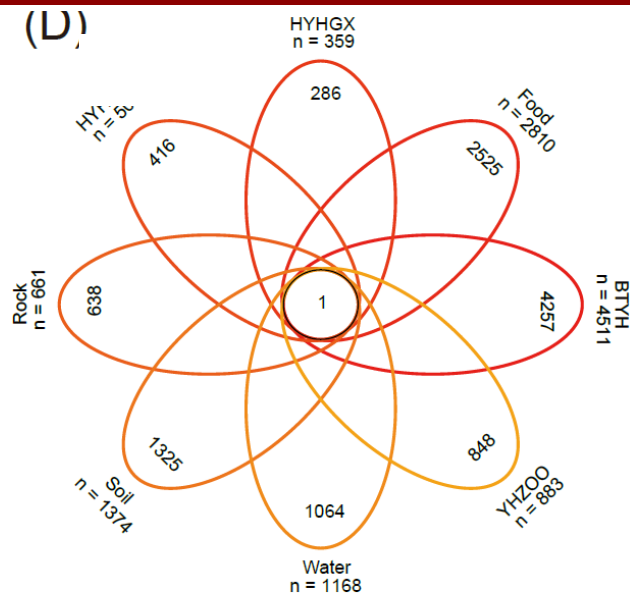


Dominant Species Composition

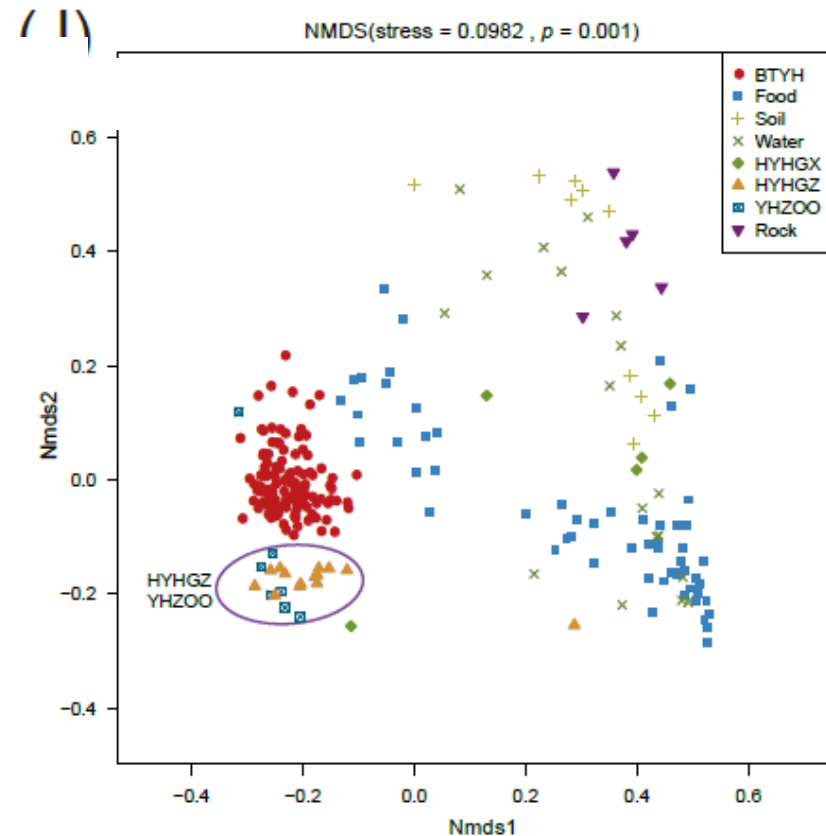




Case 2: Host environment influences gut microbial community structure



The main environmental contribution of *P. norimbergensis*



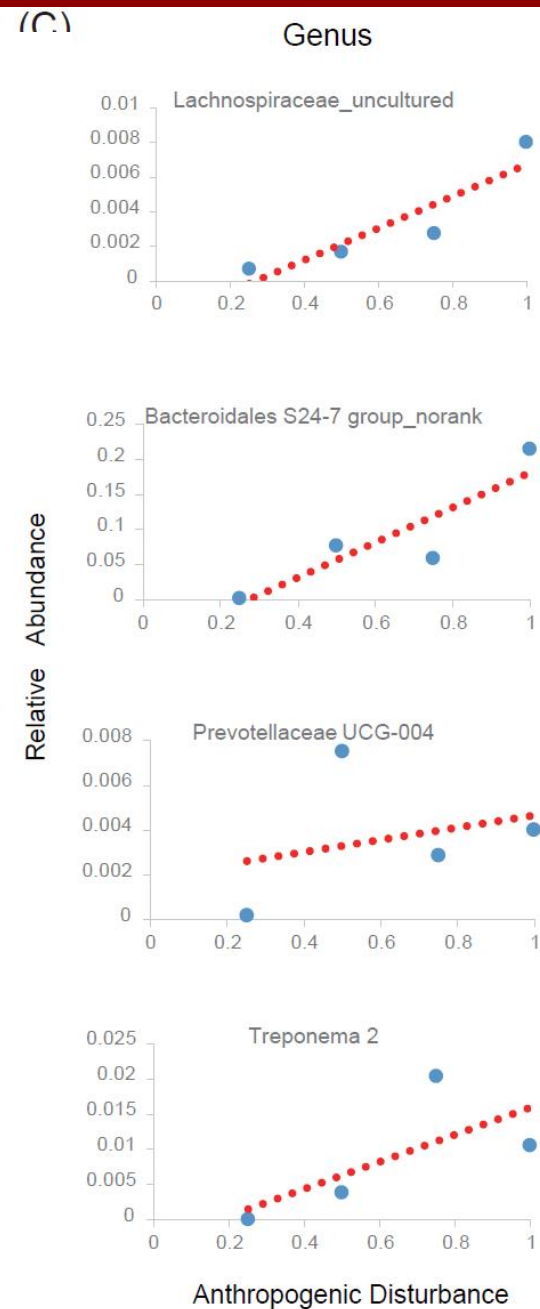
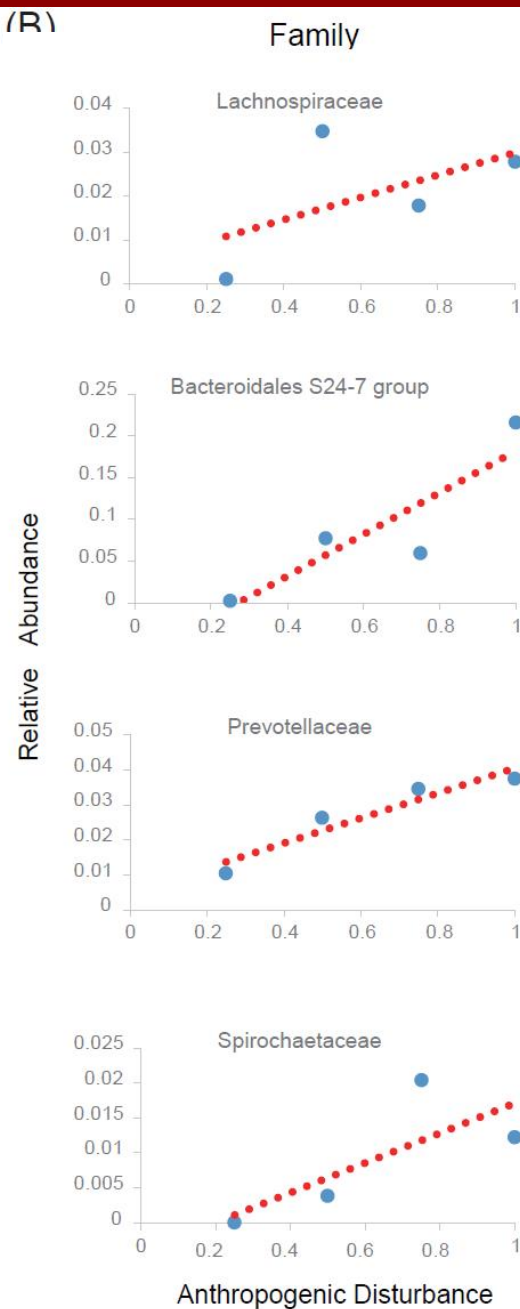
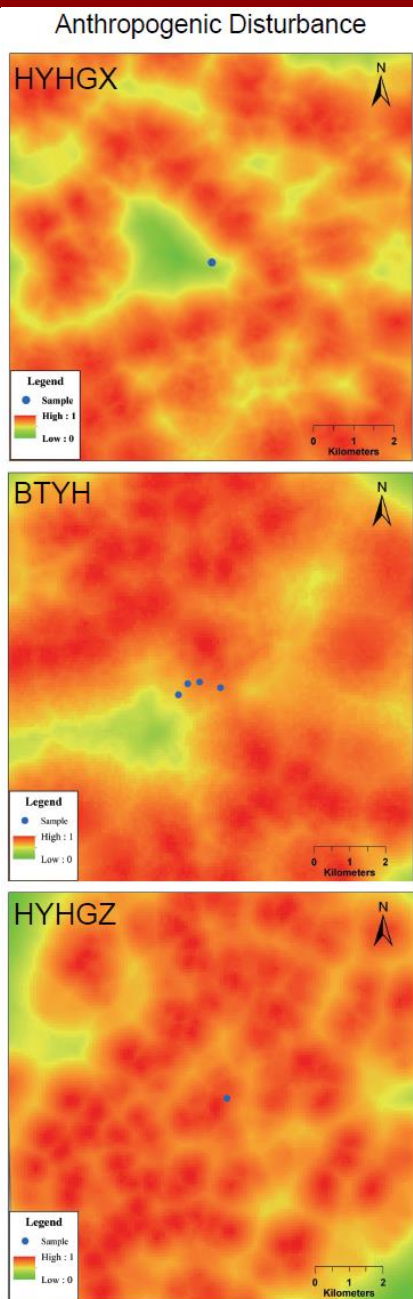
HYHGZ and YHZOO show more similar community composition



Case 3: Human disturbance drives enrichment of HDR microbes

Human disturbance gradient

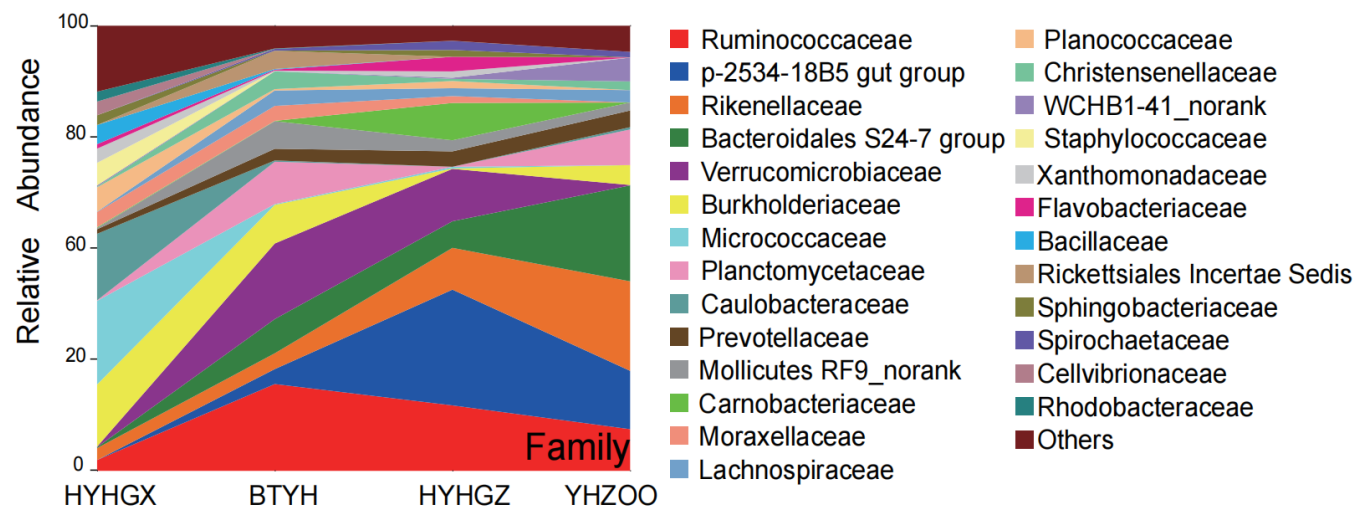
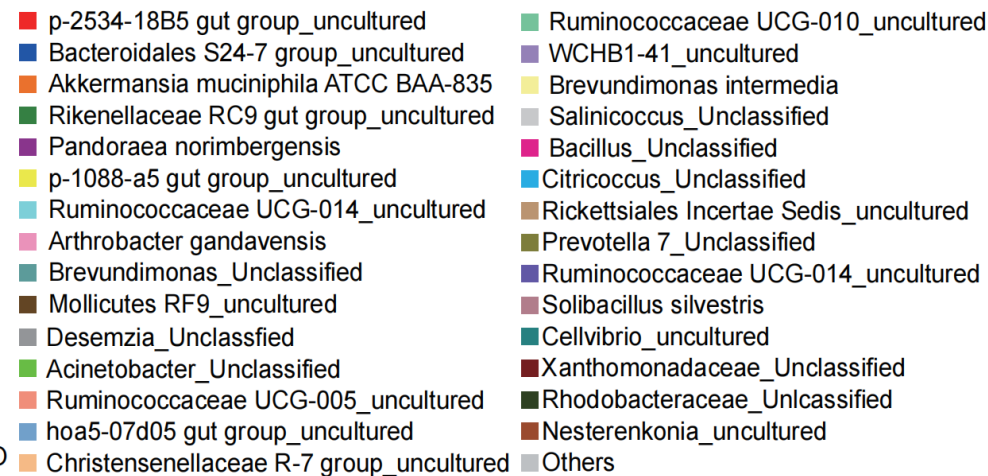
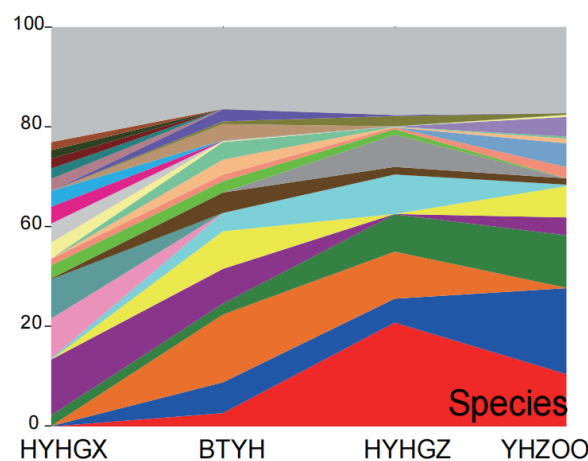
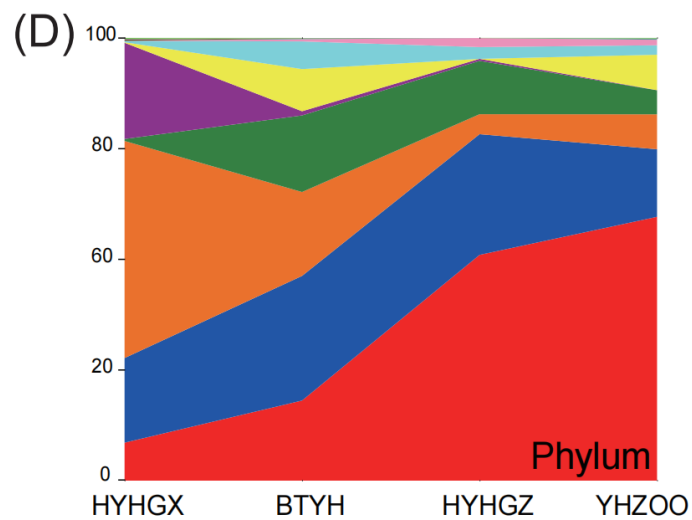
- HYHGX (Low)
- BTYH (Medium)
- HYHGZ (High)
- YHZOO (Strongest)



Human disturbance-related (HDR) gut microbial taxa increase significantly with disturbance intensity

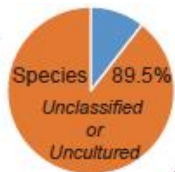


Case 3: Human disturbance drives enrichment of HDR microbes

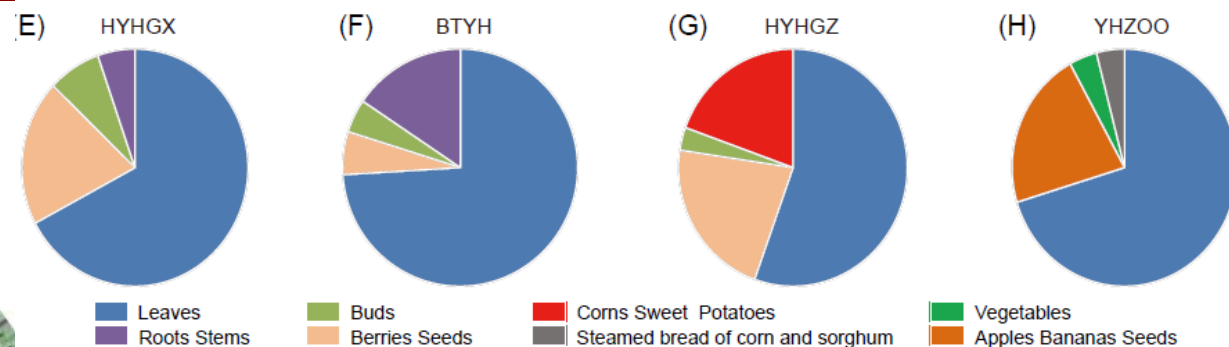
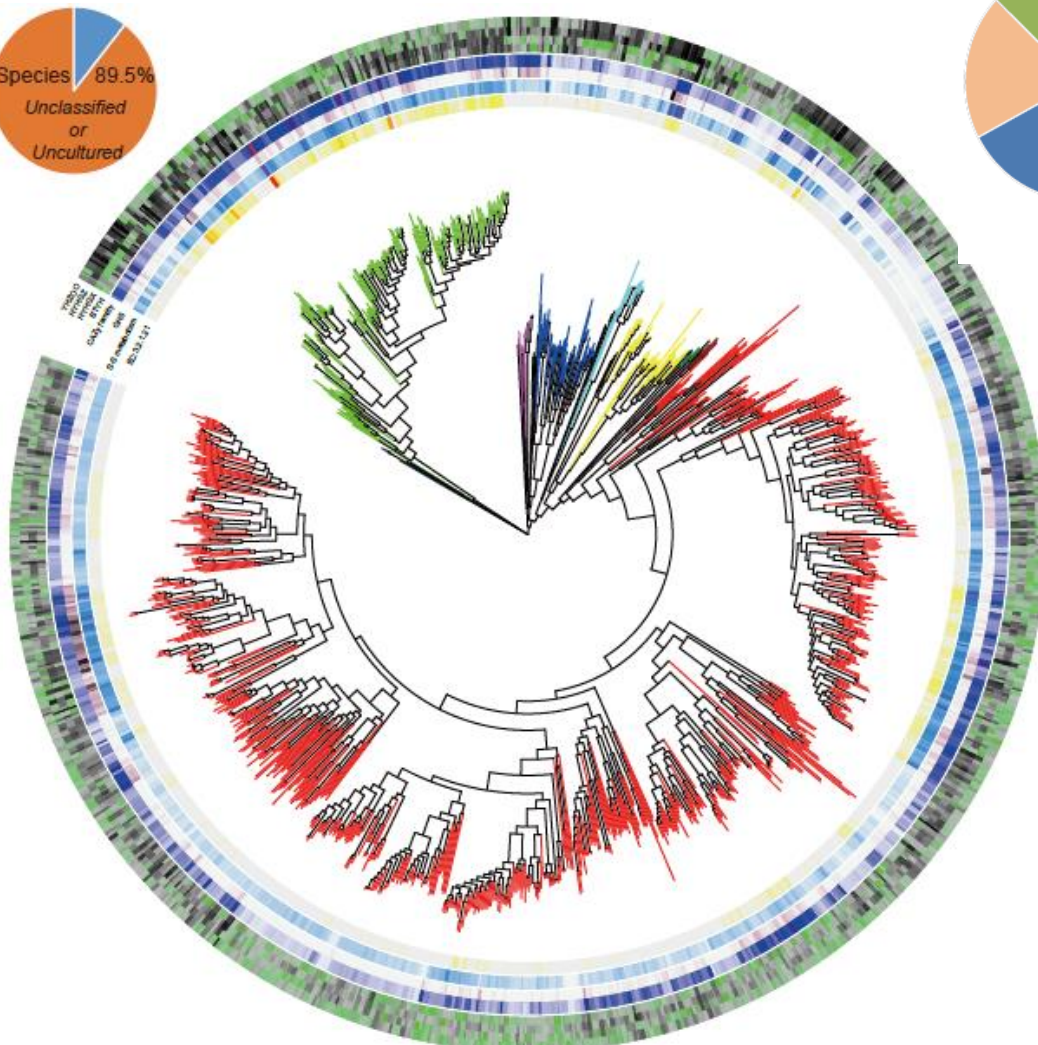
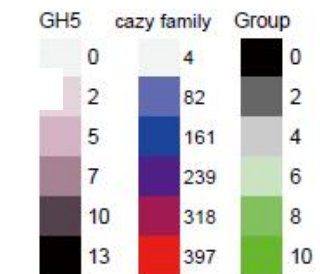
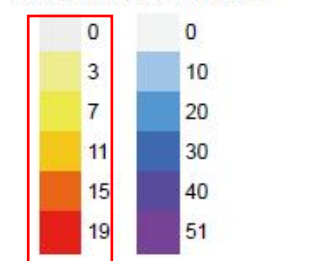


Case 4: Cellulose degradation function

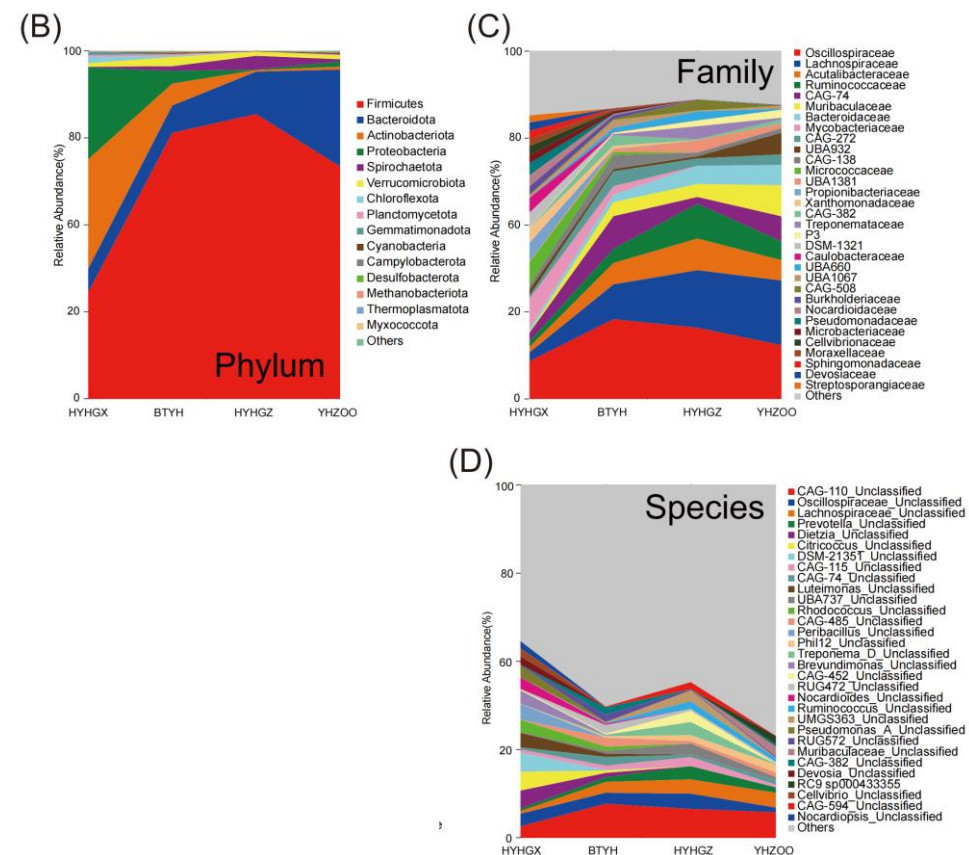
Phylum	Number of bins
Firmicutes	840
Bacteroidota	189
Proteobacteria	59
Actinobacteriota	51
Verrucomicrobiota	15
Spirochaetota	14
Thermoplasmata	4
Cyanobacteria	4
Planctomycetota	4
Others	19



EC:3.2.1.21 S-S metabolism

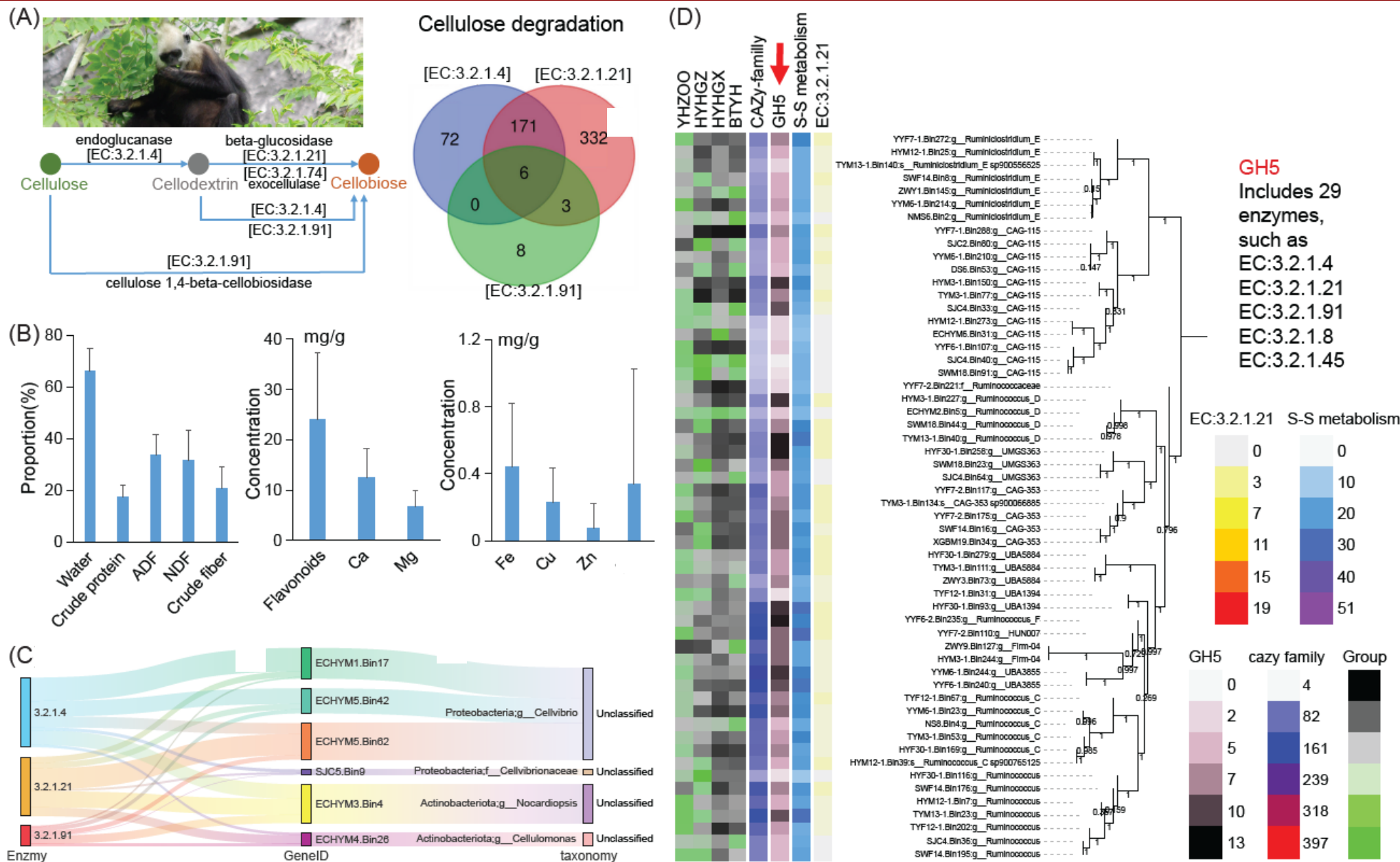


Based on feeding/feeding-record data



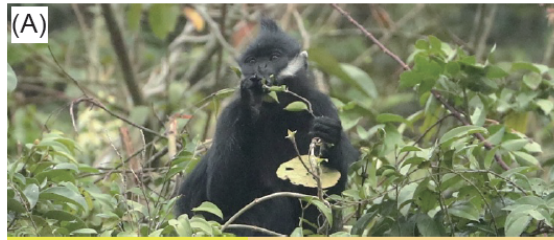
1,199 high-quality non-redundant MAGs

Case 4: Cellulose degradation function

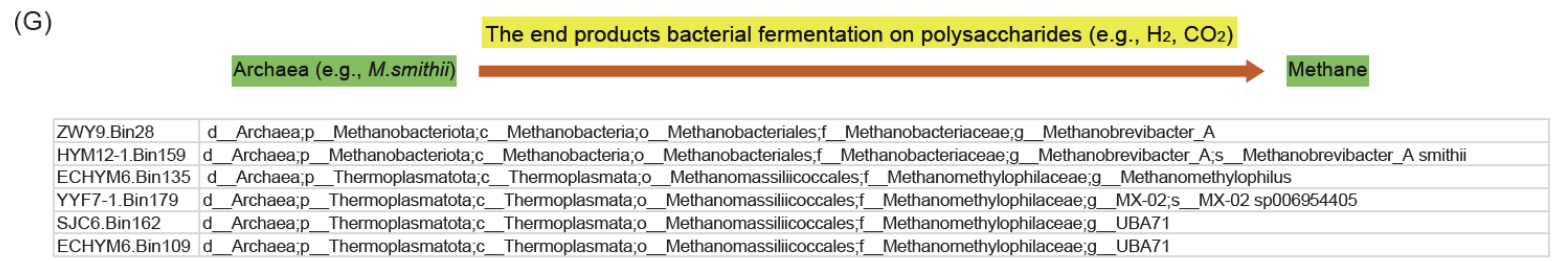
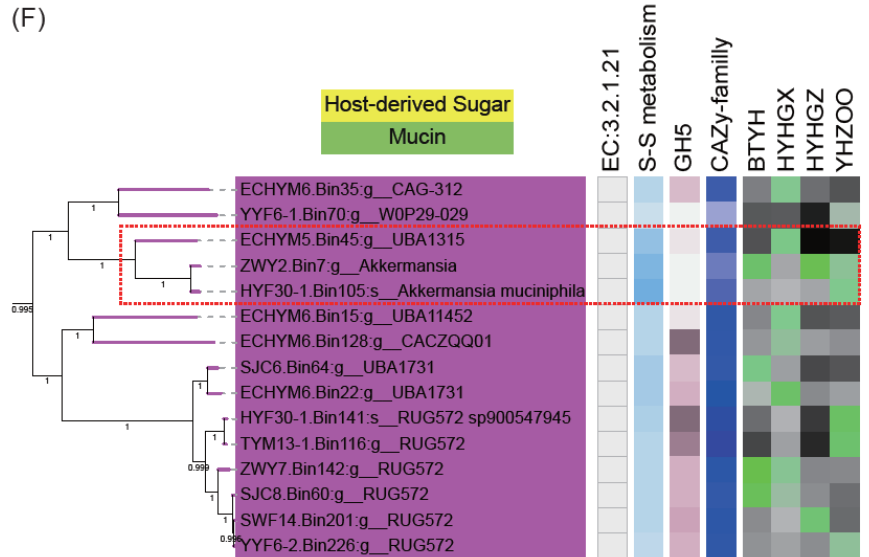
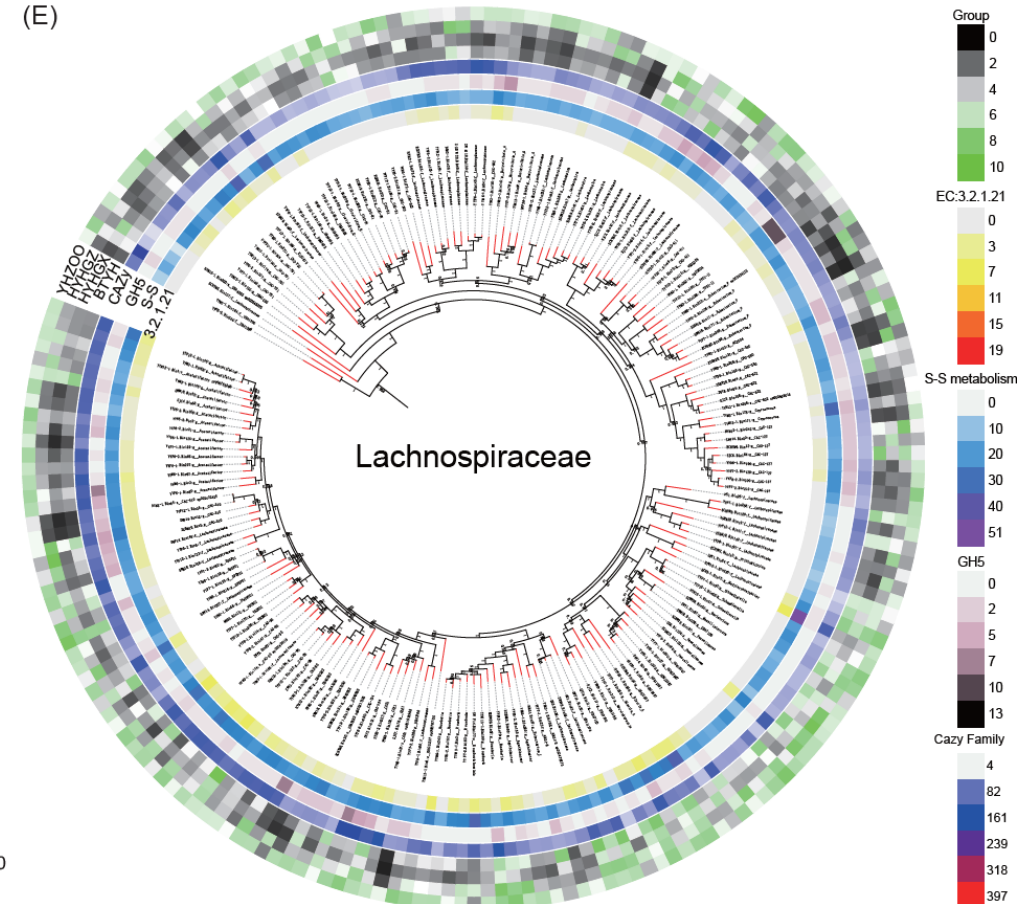
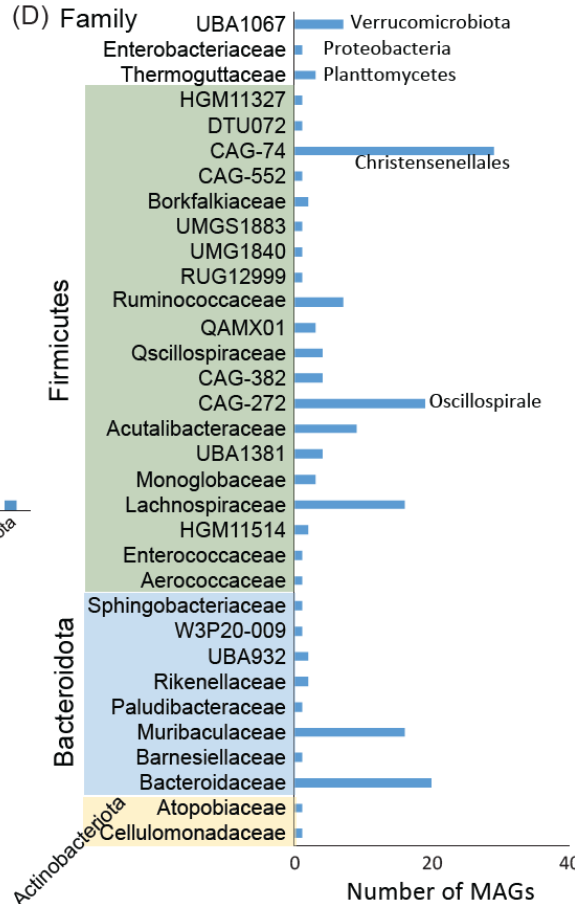
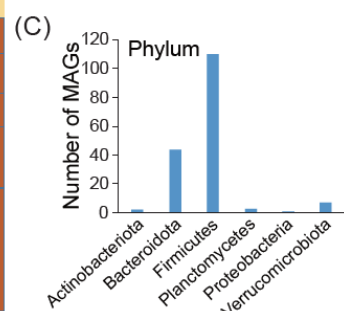
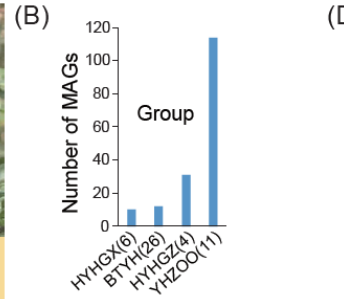


1,199 MAGs reveal potential roles in cellulose degradation

Case 5: Degradation Potential of Dietary Polysaccharides and Host Glycans



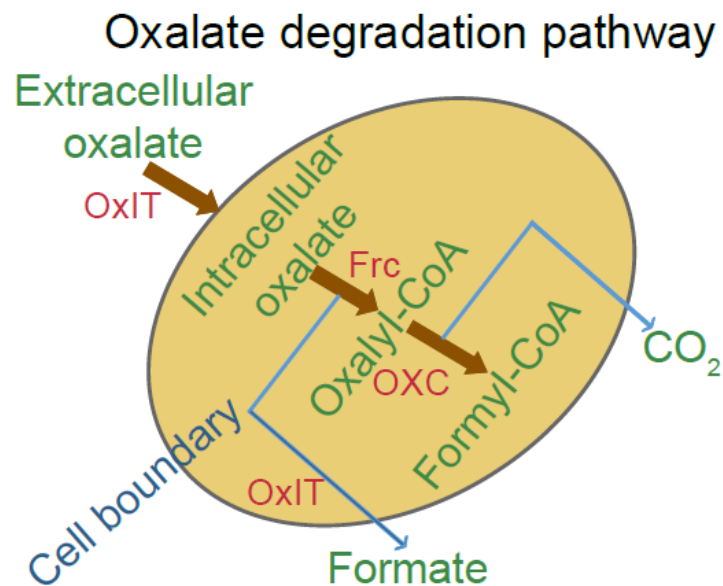
Major diet-derived polysaccharides	Microbial carbohydrate-degrading enzymes activities
Starch	GH13,GH31,GH97
Inulin	GH32,GH91
Pectin	GH28,GH8,CE12,PL1,PL9
Cellulose	GH1,GH3,GH5,GH8,GH9,GH44,GH48
Arabinoxylan	GH3,GH5,GH11,GH43,GH51,GH67,GH115,CE1,CE2,CE4,CE6,CE7,GH44,GH48



Potential Role of MAGs in Polysaccharide & Glycan Degradation

Case 6: Oxalate Degradation Function and Adaptation

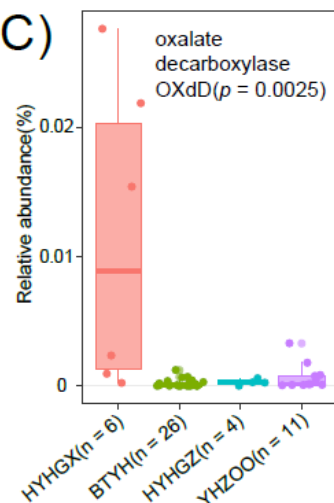
(A)



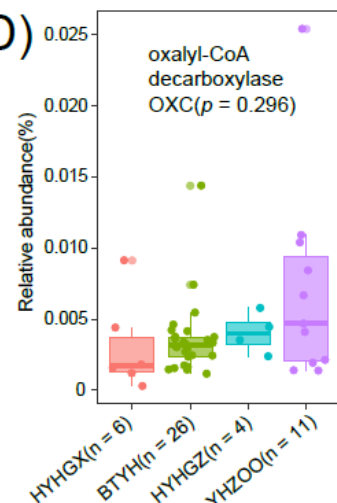
(B)

MAGs	OXdD	OXC	Group	Taxon
ECHYM4.Bin55	0	1	HYHGX	p_Actinobacteriota;c_Actinomycetia;o_Mycobacteriales;f_Mycobacteriaceae;g_Rhodococcus
ECHYM6.Bin150	1	0	HYHGX	p_Firmicutes;c_Bacilli;o_Bacillales_A;f_Planococcaceae;g_Solibacillus
ECHYM6.Bin27	1	0	HYHGX	p_Firmicutes;c_Bacilli;o_Bacillales_B;f_DSM-1321;g_Peribacillus
ECHYM5.Bin83	0	1	HYHGX	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Burkholderiaceae;g_Oxalicibacterium
ECHYM6.Bin121	0	1	HYHGX	p_Proteobacteria;c_Gammaproteobacteria;o_Enterobacterales;f_Enterobacteriaceae;g_Escherichia;s_Escherichia coli
SJC4.Bin44	0	1	BTYH	p_Actinobacteriota;c_Coriobacteriia;o_Coriobacteriales;f_Eggerthellaceae;g_Z304
ZWY3.Bin15	0	1	BTYH	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_Paramuribaculum
ZWY7.Bin134	0	1	BTYH	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_UBA7173
SJC6.Bin47	0	1	BTYH	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Burkholderiaceae;g_Oxalobacter
SJC8.Bin24	0	1	BTYH	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Burkholderiaceae;g_Oxalobacter
NMS5.Bin15	0	1	BTYH	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Burkholderiaceae;g_Oxalobacter;s_Oxalobacter sp900760095
SWF14.Bin65	0	1	HYHGX	p_Actinobacteriota;c_Coriobacteriia;o_Coriobacteriales;f_Eggerthellaceae;g_Adlercreutzia
XGBM19.Bin60	0	1	HYHGX	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-485
XGBM19.Bin68	0	1	HYHGX	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
SWF14.Bin89	0	1	HYHGX	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_Paramuribaculum
XGBW28.Bin190	0	1	HYHGX	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_Paramuribaculum
XGBM19.Bin61	0	1	HYHGX	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_UBA7173
SWM18.Bin107	0	1	HYHGX	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_P3;g_Phil12
YYF6-2.Bin188	0	1	YHZOO	p_Actinobacteriota;c_Actinomycetia;o_Actinomycetales;f_Bifidobacteriaceae
YYF7-1.Bin121	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Bacteroidaceae;g_Phocaeicola
TYF12-1.Bin81	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-485
TYM3-1.Bin25	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-485
YYM6-1.Bin90	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-485
TYM3-1.Bin201	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-485;s_CAG-485 sp002491165
HYF30-1.Bin20	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
HYF30-1.Bin227	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
HYM3-1.Bin182	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
TYF12-1.Bin29	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
TYF12-1.Bin42	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
TYF12-1.Bin75	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
TYF12-1.Bin232	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873
TYF12-1.Bin233	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873;s_CAG-873 sp900554715
HYF30-1.Bin240	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_CAG-873;s_CAG-873 sp900555715
HYF30-1.Bin281	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_Muribaculum;s_Muribaculum sp002492595
TYM13-1.Bin253	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_Paramuribaculum
TYF12-1.Bin60	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_Paramuribaculum;s_Paramuribaculum sp900759835
TYF12-1.Bin136	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_UBA3263
YYM6-1.Bin209	0	1	YHZOO	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_UBA7173
HYM12-1.Bin234	0	1	YHZOO	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Burkholderiaceae;g_Oxalobacter

(C)



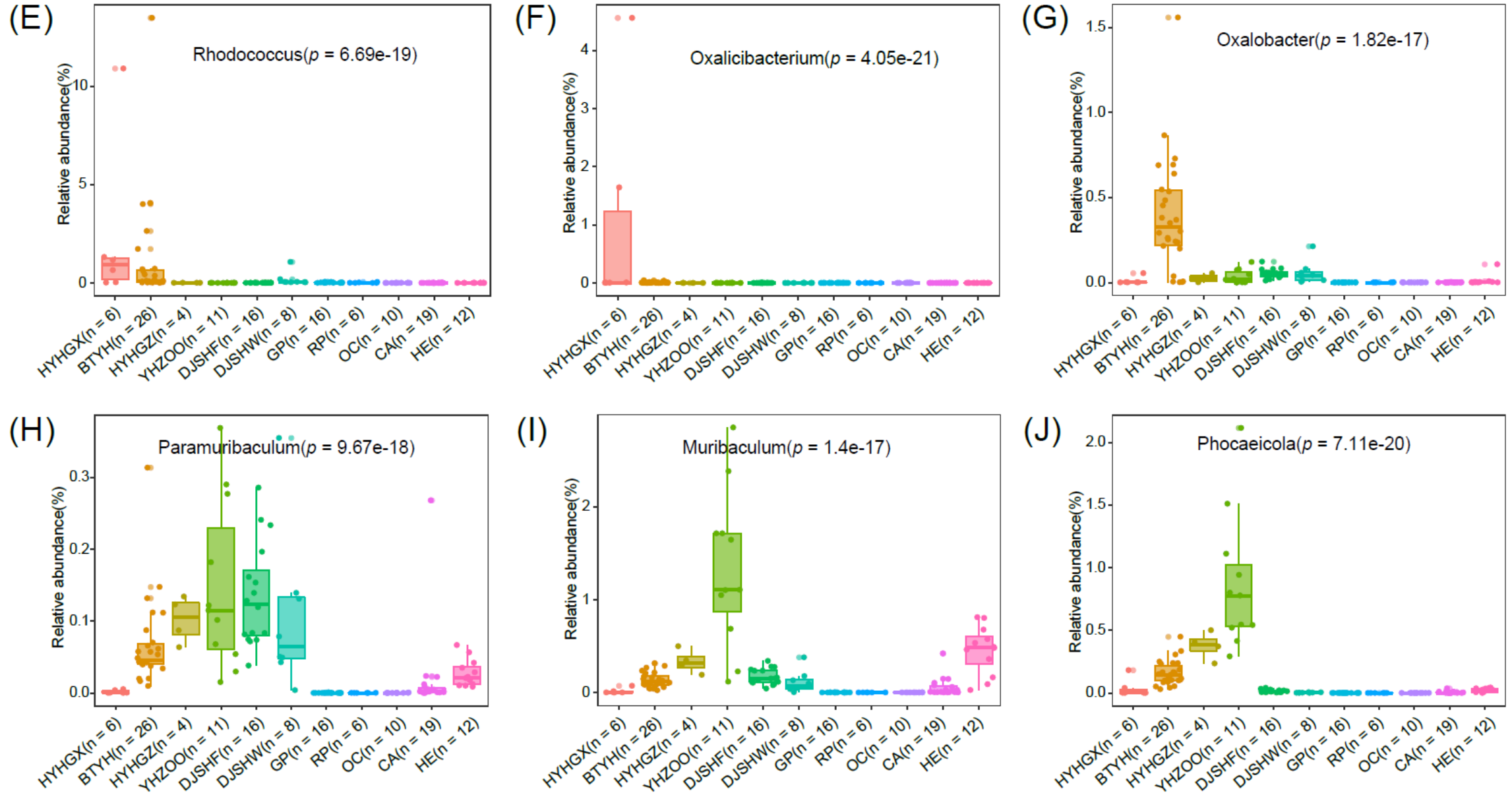
(D)



Potential Role of Langur Gut Microbiome-Associated MAGs in Oxalate Degradation



Case 6: Oxalate Degradation Function and Adaptation



Potential Role of Langur Gut Microbiome-Associated MAGs in Oxalate Degradation



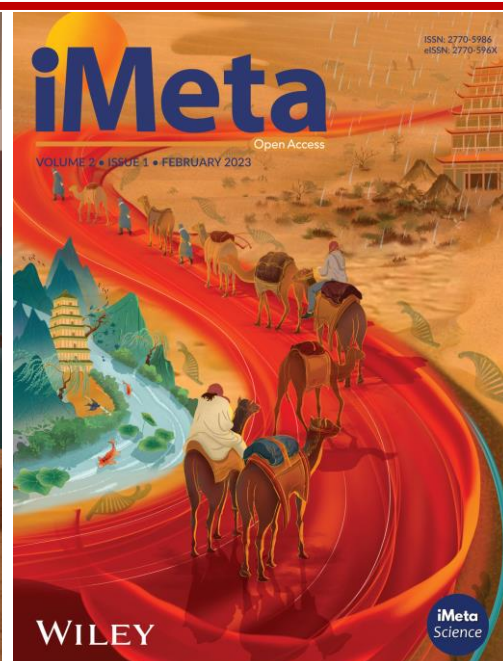
Summary

- ❑ Significant differences in gut microbial diversity and functions across langur populations.
- ❑ With increasing human disturbance, we found a significant enrichment of human disturbance-related microbes in langurs, suggesting that human activities are directly driving the “humanization” of their gut microbiota.
- ❑ The langur gut microbiome not only degrades polysaccharides but also supports oxalate metabolism and mucin utilization. These functions provide adaptive strategies that help langurs in karst limestone regions cope with high-fiber and oxalate-rich diets.

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